

Current trends in multiple sclerosis research : an update on pathogenic concepts

Joris VANDERLOCHT, Niels HELLINGS, Jerome J. A. HENDRIKS and Piet STINISSEN

Hasselt University, Biomedical Research Institute and Transnationale Universiteit Limburg, School of Life Sciences, Diepenbeek, Belgium

Abstract

Multiple sclerosis (MS) is a chronic inflammatory disease of the central nervous system (CNS) of presumed autoimmune origin which develops in a genetic susceptible individual triggered by additional environmental factors. In this review, we will provide an update of basic pathogenic concepts. In addition, we will discuss newly evolving concepts in MS pathogenesis such as pathogenic heterogeneity, importance of axonal loss and the role of CD8⁺ T lymphocytes in tissue injury. In the last part of this review we will briefly describe currently approved MS treatments and summarize some promising therapeutic approaches that are currently under evaluation.

Key words : Multiple sclerosis ; axonal injury ; pathogenic heterogeneity ; CD8 T cells.

1. Clinical features

Multiple sclerosis (MS) is one of the most frequent causes of neurological impairment in young adults affecting 0.05-0.15% of Caucasians (1). The major pathological hallmark is the presence of sclerotic lesions or 'plaques', scattered throughout the central nervous system (CNS). These lesions are characterized by selective loss of myelin and oligodendrocytes leading to impaired or complete loss of axonal conduction (2). This demyelinating process is accompanied by an inflammatory reaction with infiltrates composed mainly of T cells and macrophages (3). Although almost any CNS region can be affected, there is a preference for ventricle surrounding tissue, optic nerves, brain stem and spinal cord. The most common disease symptoms include chronic or relapsing paralysis and problems of vision, sensation, strength and coordination (4). The multiplicity of clinical deficits is suggested to reflect the variety in location, size and number of lesions (5).

The diagnosis of MS is mainly based on clinical history combined with the detection of brain abnormalities, that is, demonstrating the scatter of CNS lesions in space and time (i.e. the occurrence of a second clinical episode at a different site in the

CNS). Magnetic resonance imaging (MRI) of the brain and spinal cord is the most sensitive technique aiding the diagnosis of MS (6). Recently, an international panel published new diagnostic criteria for MS (McDonald criteria), suggesting that MRI evidence of dissemination in time and space is sufficient for the diagnosis of MS even before clinical deterioration has occurred (7).

The heterogeneity of the disease course resulted in a classification of MS in two major forms. Relapsing remitting (RR-) MS is the most frequent form (80-90%) and affects women twice as often as men. This form is characterized by a relapsing remitting illness with episodes of neurological dysfunction lasting several weeks, followed by substantial or complete recovery. Many years after onset, the majority of relapsing remitting MS patients (RR-MS) develops a gradual clinical progression with or without clinical attacks superimposed. This is termed secondary progressive MS (SP-MS). A minority of patients (10-20%) display a primary progressive (PP-) MS, characterized by a gradual decline in neurological function and less evident inflammation on MRI (8). Heterogeneity of MS is not confined to disease course and clinical presentation, e.g. which areas are primarily affected. Morphological studies with MRI (9) and histopathological evaluation (10, 11) revealed profound differences among MS patients. The factors underlying this heterogeneity are not completely understood but include a complex genetic trait that translates into different immune abnormalities, increased vulnerability of CNS tissue to inflammatory insult or reduced remyelinating capacity (12).

Financial support : 'Nationaal fonds voor wetenschappelijk onderzoek Vlaanderen (FWO)'
'Bijzonder onderzoeksfonds' Hasselt University
The Belgian WOMS foundation
The Belgian Charcot foundation
Transnational university Limburg.

2. Role of genes and environment

The aetiology of MS remains unclear, but according to current data the disease develops in a genetic susceptible individual and requires additional environmental triggers (12). To establish a genetic component in a complex disease, familial aggregation studies serve as starting point. Population and family studies show that the prevalence of the disease is substantially increased in family members of MS patients with first degree relatives and daughters of affected mothers having the highest risk (13, 14). Evidence originating from twin, adoption and half-sibling studies indicates that the familial segregation of MS is related to genetic sharing rather than to shared family environment. This evidence includes the difference in concordance rates between monozygotic (20-35%) and dizygotic (5%) twins (15, 16), the similar recurrence risk between adoptive relatives of MS patients and the general population (17) and the lower occurrence risk for half-siblings (1.32%) compared to full-siblings (3.46%) (18). However, the quantitative genetic trait of MS has shown to be difficult to dissect. The only region consistently found to be associated with MS prevalence is the MHC region. The MHC locus is a four mega-base region containing the classical HLA genes as well as at least 128 other genes (19). Within this region the HLA-DR2 (DRB1*1501, DQA1*0102 and DQB1*0602) haplotype was consistently shown to be associated with MS (20). In addition, MHC class I alleles may also contribute to disease susceptibility, although associations were much weaker (21). Current studies indicate that no single major susceptibility gene exists for MS and instead many loci are believed to confer overall MS susceptibility. However, genetic predisposition alone is not sufficient to develop MS, since other interacting factors including immunological and environmental factors have been shown to contribute to disease susceptibility.

Several lines of evidence support the contribution of environmental factors to the aetiology of MS. These include the low concordance rate of identical twins (22, 23), the increased prevalence of MS with distance from the equator (16, 24) and studies showing that migration before puberty from a high to a low prevalence area results in a reduction in the risk to develop MS (25). The fact that first-degree adopted relatives of MS patients do not display an increased MS prevalence compared to the general population strongly suggests that MS is not purely a transmissible disease (17). Therefore, any environmental factor is likely to be ubiquitous and act on a population basis rather than within the family microenvironment (22). Both lifestyle influences and infectious agents have been proposed as risk factors (26). Substantial efforts were undertaken to identify an MS triggering virus (27, 28).

Although several viruses, including human herpes virus, 'MS-associated retrovirus' (MSRV), Epstein Barr virus, rabies and measles, have been claimed as potential candidate, none of the studied viruses could be univocally linked to disease (27, 28).

The lack of hard proof for an MS causing virus does not rule out the possibility that infectious triggers are involved in disease initiation. According to the 'hit and run' hypothesis, persistence of an encephalitogenic virus is not necessary for the continuation of the disease process (29). It is possible that a transient viral CNS infection causes minor damage to myelin leading to the release of myelin epitopes and subsequent activation of myelin-reactive T cells (30, 31). Alternatively, it is possible that autoreactive T cells become activated in the periphery after recognition of cross-reactive viral epitopes that mimic self peptides ('molecular mimicry') (32). Moreover, the profound heterogeneity of MS could reflect different triggering factors and thus explain why MS patients do not show universal positivity for a particular microorganism (33).

3. Multiple Sclerosis : a CNS specific autoimmune disease

Various lines of evidence suggest that MS is an autoimmune disease mediated by peripheral myelin reactive T cells (34, 35). Indirect evidence for the possible autoimmune nature stems from the animal model experimental autoimmune encephalomyelitis (EAE). This experimental disease shares many clinical and histological features with MS and is induced by generating T cell-mediated immunoreactivity to CNS antigens (36). Adoptive transfer of myelin reactive T cells to naive recipients also induces EAE, demonstrating the T cell mediated autoimmune nature of this model (37, 38). There is also direct evidence supporting MS as an autoimmune disease. For example, myelin reactive T cells accumulate in lesions and cerebrospinal fluid (CSF) of MS patients but not in the CNS of patients with other neurological diseases (39, 40). The linkage of HLA class II genes such as HLA-DR2 to disease is another important argument, because HLA-DR2 determines the ability to recognize certain epitopes of potential autoantigens (41-43).

Although the exact event leading to the activation of myelin reactive T cells remains elusive, there are several mechanisms by which an infectious trigger may induce an autoimmune disease. The main postulated mechanism is molecular mimicry (32, 44, 45) in which autoreactive T cells are activated by a viral or bacterial epitope with structural or sequential similarity to the self peptide (46, 47). Another proposed mechanism is bystander activation including both antigen-dependent and antigen-independent mechanisms (12). Antigen-independent mechanisms include activation of autoreactive T cells by

bacterial or viral superantigens (48), Toll like receptor (TLR) triggering (49) and exposure to high concentrations of cytokines secreted during unrelated immune responses (50). Moreover, antigen-dependent activation can occur when myelin epitopes are released as a result of initial damage to the CNS, as illustrated in the Theiler virus-induced EAE model (30, 51). In contrast to the above mentioned mechanisms that suggest activation in the periphery, recent evidence in several animal models for MS reveals that activation of autoreactive T cells can also be initiated in the CNS by a discrete population of vessel-associated dendritic cells (52, 53). In this light, it was recently shown that besides activated T cells also naive T cells cross the blood brain barrier (BBB), bypassing the need for antigen priming in the periphery (52-54). This is probably facilitated by the increased expression of adhesion ligands (ICAM-1 and VCAM) during CNS inflammation in EAE (55).

Once activated, autoreactive T cells expand and traffic to the CNS (56). Migration through the endothelium of the BBB is facilitated by the expression of adhesion molecules, chemokines, their receptors and the release of pro-inflammatory cytokines (57-60). Once autoreactive T cells have extravasated through the endothelium, they have to pass through a barrier of extracellular matrix to enter the CNS. This is mediated by the secretion of matrix metalloproteases (MMP-2 and MMP-9) following contact with collagen. These enzymes are not only involved in the proteolytic cleaving of extracellular matrix but can also cleave myelin components and thus generate immunogenic peptides (61-63). Thus, beside their role in opening the blood brain barrier these MMPs are also able to perpetuate inflammatory responses.

Within the CNS, T cells become reactivated when they encounter their specific myelin epitope presented by resident antigen presenting cells (APC). The nature of the APC responsible for antigen presentation is still subject of debate. Although astrocytes (64) and microglia (65-67) are capable of presenting antigen, recent evidence in EAE suggests that antigen presentation by perivascular dendritic cells and macrophages is sufficient to develop disease (53). Following reactivation, autoreactive T cells spread into the white matter and produce pro-inflammatory cytokines which trigger a cascade of immune reactions. During this inflammatory response, chemokines are released and the expression of adhesion molecules on endothelial cells is increased. This results in the recruitment of other immune cells including monocytes, T cells, mast cells and B cells. These inflammatory events ultimately lead to demyelination, astrogliosis and loss of oligodendrocytes and neurons. This tissue damage is induced by the combined effects of oxygen radicals, cytokines, autoantibodies, cytotoxic cells (CD8⁺ T cells, $\gamma\delta$ T cells and macrophages),

complement deposition and myelin phagocytosis (68, 69). The decline of this inflammatory event is paralleled with the clearance of debris by macrophages and a relative dominance of Th2 cytokines. Surviving oligodendrocytes and oligodendrocyte precursors are activated and spontaneous myelin repair occurs in about 40% of plaques (70-72). Despite the fact that remyelination is wide spread, it is also clear that much of the myelin damage is not repaired (73). Especially lesions in patients with long standing disease display less pronounced remyelination (74, 75). This indicates that recurrent episodes of inflammation may exhaust the ability of oligodendrocytes to regenerate myelin (71) or lead to a milieu that is no longer favourable for remyelination (76). One of the challenges in MS research is to develop strategies that enhance inherent repair mechanisms, while simultaneously limiting immune-mediated damage.

4. New concepts in MS research

The view that MS is as a CD4⁺ T cell driven demyelinating autoimmune disease is too simplistic. Indeed, accumulating evidence suggests a possible pathogenic role for CD8⁺ T cells and besides demyelination also axonal degeneration is a prominent pathological feature. In addition, recent insights reveal a profound pathological heterogeneity among MS patients. These novel directions in MS research will be addressed in the next paragraphs.

4.1. PATHOLOGICAL HETEROGENEITY

Recent evidence suggests that distinct pathological subtypes may exist in MS (77). The analyses of a large number of MS lesions in the active stage of demyelination revealed at least four distinct pathological patterns. These patterns differ in the extent of myelin and oligodendrocyte loss, the degree of humoral contribution (complement and antibody deposition) and the involvement of different immune cells. Pattern I and II share a number of similarities, such as prominent T cell and macrophage infiltration, location of lesions around venules and the occurrence of remyelination. Pattern II is distinguished from I by a prominent humoral component with deposition of antibodies and complement proteins. Based on parallels with focal cerebral ischemia, pattern III lesions are believed to be caused by a vasculitic mechanism. These lesions are characterized by a preferential loss of MAG (myelin-associated glycoprotein), oligodendrocyte apoptosis and conservation of a rim of myelin surrounding venules. Pattern IV involves non-apoptotic oligodendroglial cell death. This pathological subtype has only been found in a small subset of patients with a primary progressive

disease course (11). While in lesions of pattern I and II, the myelin sheath is the main target of the destructive process, patterns III and IV show primarily oligodendrocyte loss (78). It remains to be established whether these lesional patterns are constant along disease progression and whether these subtypes can be used for prognosis. Interestingly, different acute demyelinating lesions within one patient showed a similar pattern, indicating that lesion heterogeneity does not reflect different lesional stages, but rather distinct pathological mechanisms.

Subdividing MS patients based on their disease mechanism holds promise for more specific therapies. However, it remains to be established whether subdividing MS patients based on their lesional pattern is sufficient. Additional sub-classifications based on remyelinating capacity, responsiveness to treatment, amount of atrophy and other criteria may be needed. The identification of new MRI criteria (79, 80), CSF markers (81) and molecular markers associated with heterogeneity, will allow tailoring specific treatments for a specific subgroup of MS patients.

4.2. AXONAL PATHOLOGY

Historically, MS has been considered an inflammatory demyelinating disease with a relative preservation of axons (82). Accordingly, research focused primarily on the inflammatory response and loss of myelin. New insights regarding the timing, extent and consequences of axonal loss in MS led to renewed attention for neurodegenerative processes. Axonal ovoids indicating recently transected axons and APP (amyloid precursor protein) positive neurons reflecting impaired axonal transport can be detected in lesions of patients with a short disease duration (83). Subsequent morphologic studies confirmed that axonal transection is equally prominent in active lesions of patients with short disease duration and correlates with inflammatory activity in these lesions (84-86). In addition, acute axonal injury was also detected in inactive lesions (85). This is not observed in remyelinated lesions indicating that the myelin sheath itself may protect the axon against proinflammatory mediators released within the plaques (78) and oligodendrocytes may provide trophic support for the axon. The mechanisms of early axonal injury in MS are poorly understood, but may include deleterious effects of inflammatory mediators, T cells, oedema, glutamate, nitric oxide and genes involved in axonal responses to inflammation (87). This early axonal loss remains clinically silent for many years, suggesting that the CNS can compensate for neuronal loss (88). Irreversibly neurologic disability develops when a threshold of axonal loss is reached and the CNS compensatory mechanisms are exhausted (88-90).

4.3. A PATHOGENIC ROLE FOR CD8⁺ T CELLS

MS is generally considered to be a CD4⁺ Th1-mediated autoimmune disease (91). This view is based on the close similarities in pathology between MS and EAE (36, 37, 92). The role of CD4⁺ T cells in MS is further supported by the association of MHC class II genes with the disease and the expression pattern of chemokines, cytokines and their receptors in MS lesions and CSF that is consistent with a CD4⁺ Th1-mediated immune response (93-95). Other observations suggest an important role for CD8⁺ T cells in the pathogenesis of MS. CD8⁺ T cells have been shown to predominate in early MS lesions suggesting that their presence is not due to the recruitment of non-specific immune cells once inflammation has started (96, 97). Clonal expansion was more frequently detected among CD8⁺ T cells as compared to CD4⁺ T cells in CSF of MS patients. In addition, increased numbers of memory CD8⁺ T cells with evidence for clonal expansion have been observed in the CSF and blood of MS patients as compared to controls (98, 99). In the context of effector functions, CD8⁺ T cells are better equipped to mediate direct CNS damage. Indeed, MHC class I molecules are, unlike MHC II molecules, expressed on CNS cells such as neurons, oligodendrocytes, axons and astrocytes (100, 101). In addition, a number of HLA class I-restricted myelin epitopes have been described for myelin basic protein (MBP), proteolipid protein (PLP) and MAG (102-105). Myelin specific CD8⁺ T cells are able to induce severe EAE upon adoptive transfer (106, 107). Moreover, myelin reactive CD8⁺ T cells have been isolated from MS patients and were able to specifically lyse oligodendrocytes and myelin pulsed target cells in vitro (100, 103, 105, 108). CD8⁺ T cells have also been found in MS lesions in proximity of axons and oligodendrocytes with their cytotoxic granules polarized towards the CNS target cells (109, 110). Since there is no information on the antigen specificity of the infiltrated CD8⁺ T cells it cannot be ruled out that these cells may be regulatory rather than pathogenic. Anti-idiotypic CD8⁺ T cells for instance are part of the regulatory network that is thought to control autoreactive T cells by recognition of clonotypic determinants (111).

5. MS therapies : established treatments and new hopes

The increased understanding of the MS pathogenesis has led to the implementation of a number of immunotherapeutic approaches. The currently approved treatments and some promising therapeutic approaches that are currently under evaluation will be discussed below.

5.1. ESTABLISHED MS THERAPIES

Interferon- β and glatiramer acetate are drugs currently approved for the treatment of RR-MS. IFN- β is a naturally occurring cytokine and is the most broadly used drug in MS therapy. Three preparations of IFN- β (IFN- β 1a : AvonexTM, RebifTM and IFN- β 1b : BetaseronTM) have shown efficacy in reducing the relapse rate and the number of active lesions in RR-MS (112-115). The predominant mechanism of action is unknown, but may involve the suppression of T cell proliferation and shifting of the T cell cytokine secretion from a pro-inflammatory Th1 profile towards a more protective Th2 profile (116). Other immunomodulatory activities include the induction of IL-10 and neurotrophic factors, blocking of BBB opening via inhibition of MMP-2 and -9 and reduction of cell adhesion to the BBB (12).

Glatiramer acetate (Co-polymer 1 : CopaxoneTM) is a random polymer of 4 amino acids (L-glutamic acid, L-lysine, L-alanine and L-tyrosine) that mimics MBP. Its main mechanism of action involves the induction of a Th1-Th2 shift. Other activities of GA include the induction of anergy of MBP-reactive T cells, cross-reactivity with myelin epitopes, polyclonal activation of T cells leading to bystander suppression and induction of neurotrophic factors such as BDNF (117-120).

However, treatment with GA or IFN- β has substantial limitations. Although the annual relapse rate is decreased by about one third (121-123), their long-term clinical effect is uncertain (124). Recent papers report negligible effects of the above treatments on disability in chronic stages of the disease, implying that their effects on axonal damage are probably very limited (113, 125). Available data on IFN- β treatment suggest that it might be more effective when given early, maybe even at the first clinical presentation of the disease ; long-term data are awaited (126-128).

Mitoxantrone is an anthracyclin-based chemotherapeutic that is approved for the treatment of secondary progressive and progressive relapsing MS. Treatment with mitoxantrone resulted in significantly fewer lesions, reduced relapse rate and reduced progression of disability (129, 130). Proposed mechanisms of action are suppressing T and B cell responses, but also inducing apoptosis in antigen presenting cells and macrophages (131).

Acute exacerbations of MS are generally treated with high doses of glucocorticoids, such as methylprednisolone (132-134). The above described approved MS therapies are all directed at suppressing or modulating immune responses. They are only partially effective in controlling MS and were introduced before there was an understanding of their mode of action. Elucidation of the mechanisms by which these therapeutics exert their protective effects and further insights in pathogenic

pathways of MS have led to new therapeutic strategies.

5.2. NEW THERAPEUTIC APPROACHES

Future immunotherapies can be classified into 3 broad categories (135). The first are antigen-specific therapies. These include T cell vaccination (111, 136), T cell receptor (TCR) vaccination (137), oral tolerance, altered peptide ligands (91) and MHC blockers. Importantly, these approaches require detailed knowledge of all involved autoantigens and effector cells. A second class of novel immunotherapies relies on targeting well defined pathogenic mechanisms. Promising examples of these approaches are daclizumab, a monoclonal antibody specific for the subunit of the interleukin-2 receptor influencing activation of peripheral CD4⁺ T cells (138) ; natalizumab, a monoclonal humanized antibody directed against VLA-4 inhibiting cell adhesion to the BBB (139), and modulators of cAMP levels in the brain and in immune cells (140). The last class of new immunotherapies are agents with broad immunomodulatory activities. These include for example alemtuzumab, a humanized monoclonal antibody directed against CD52 that is intended to deplete T and B cells, monocytes and macrophages (141), statins, currently approved cholesterol lowering agents with immunomodulatory properties (142-144), pregnancy related hormones (145) and haematopoietic stem cell transplantation.

Besides the immunologic aspects, attention has in recent years focused on the enhancement of repair and regeneration mechanisms as targets for therapy especially in secondary progressive MS. There are two broad approaches for promoting myelin repair (76). Transplanting cells with myelinogenic capacity represents the first approach. This approach has made significant progress in animal models for MS (146-148). The second approach is directed at promoting the endogenous protective pathways from the brain and immune system. For this approach, a number of strategies have shown an effect in animal models (149) and await validation in MS.

All therapeutics summarized here are in different stages of development. While one important goal is to develop new therapeutics, there is also a growing appreciation that some MS therapeutics may be beneficial when given in combination (150). The prerequisite of combination therapy is that both drugs have an additive or synergistic effect, but not overlapping toxicities (151).

6. Summary

Multiple sclerosis (MS) is generally considered a CD4⁺ T cell-mediated autoimmune disease that is

triggered by unknown exogenous agents in subjects with a specific genetic background. New tools, such as gene expression profiling with cDNA microarrays and LD-based maps may help us identify disease-risk alleles. Despite major efforts, the precise cause of MS remains unknown and many aspects of MS pathogenesis have become more complicated. Whereas demyelination was originally thought to be relevant for the lasting neurological deficit, it is now commonly accepted that the extent of axonal loss dictates the degree of permanent clinical disability. How axonal damage can be prevented remains elusive. In addition, new insights on the prominent role of CD8⁺ T cells indicate that the concept of MS as CD4⁺ driven autoimmune disease should be revisited. Finally, different patterns of tissue damage have been shown in active MS lesions, suggesting that the mechanisms of injury are distinct in different subgroups of patients. The classification of pathogenic mechanisms in an individual patient may be necessary to provide better targeted therapies.

REFERENCES

1. NOSEWORTHY J. H., LUCCHINETTI C., RODRIGUEZ M., WEINSHENKER B. G. Multiple sclerosis. *N. Engl. J. Med.*, 2000, **343** (13) : 938-952.
2. HICKEY W. F. The pathology of multiple sclerosis : a historical perspective. *J. Neuroimmunol.*, 1999, **98** (1) : 37-44.
3. LASSMANN H., BRUCK W., LUCCHINETTI C. Heterogeneity of multiple sclerosis pathogenesis : implications for diagnosis and therapy. *Trends Mol. Med.*, 2001, **7** (3) : 115-121.
4. MCFARLIN D. E., MCFARLAND H. F. Multiple sclerosis (first of two parts). *N. Engl. J. Med.*, 1982, **307** (19) : 1183-1188.
5. SMITH K. J., McDONALD W. I. The pathophysiology of multiple sclerosis : the mechanisms underlying the production of symptoms and the natural history of the disease. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*, 1999, **354** (1390) : 1649-1673.
6. FAZEKAS F., BARKHOF F., FILIPPI M., GROSSMAN R. I., LI D. K., McDONALD W. I., MCFARLAND H. F., PATY D. W., SIMON J. H., WOLINSKY J. S., MILLER D. H. The contribution of magnetic resonance imaging to the diagnosis of multiple sclerosis. *Neurology*, 1999, **53** (3) : 448-456.
7. McDONALD W. I., COMPSTON A., EDAN G., GOODKIN D., HARTUNG H. P., LUBLIN F. D., MCFARLAND H. F., PATY D. W., POLMAN C. H., REINGOLD S. C., SANDBERG-WOLLHEIM M., SIBLEY W., THOMPSON A., VAN DEN NOORT S., WEINSHENKER B. Y., WOLINSKY J. S. Recommended diagnostic criteria for multiple sclerosis : Guidelines from the International Panel on the Diagnosis of Multiple Sclerosis. *Annals of Neurology*, 2001, **50** (1) : 121-127.
8. THOMPSON A. J., POLMAN C. H., MILLER D. H., McDONALD W. I., BROCHET B., FILIPPI M., X., DE SA J. Primary progressive multiple sclerosis. *Brain*, 1997, **120** (Pt 6) : 1085-1096.
9. MCFARLAND H. F. Correlation between MR and clinical findings of disease activity in multiple sclerosis. *AJNR Am. J. Neuroradiol.*, 1999, **20** (10) : 1777-1778.
10. RAINE C. S., SCHEINBERG L. C. On the immunopathology of plaque development and repair in multiple sclerosis. *J. Neuroimmunol.*, 1988, **20** (2-3) : 189-201.
11. LUCCHINETTI C., BRUCK W., PARISI J., SCHEITHAUER B., RODRIGUEZ M., LASSMANN H. Heterogeneity of multiple sclerosis lesions : implications for the pathogenesis of demyelination. *Ann. Neurol.*, 2000, **47** (6) : 707-717.
12. SOSPEDRA M., MARTIN R. Immunology of multiple sclerosis. *Annu. Rev. Immunol.*, 2005, **23** : 683-747.
13. SADOVNICK A. D., BULMAN D., EBERS G. C. Parent-child concordance in multiple sclerosis. *Ann. Neurol.*, 1991, **29** (3) : 252-255.
14. EBERS G. C., KOOPMAN W. J., HADER W., SADOVNICK A. D., KREMENCHUTZKY M., MANDALFINO P., WINGERCHUK D. M., BASKERVILLE J., RICE G. P. The natural history of multiple sclerosis : a geographically based study : 8 : familial multiple sclerosis. *Brain*, 2000, **123** Pt 3 : 641-649.
15. EBERS G. C., BULMAN D. E., SADOVNICK A. D., PATY D. W., WARREN S., HADER W., MURRAY T. J., SELAND T. P., DUQUETTE P., GREY T. A population-based study of multiple sclerosis in twins. *N. Engl. J. Med.*, 1986, **315** (26) : 1638-1642.
16. SADOVNICK A. D., ARMSTRONG H., RICE G. P., BULMAN D., HASHIMOTO L., PATY D. W., HASHIMOTO S. A., WARREN S., HADER W., MURRAY T. J. A population-based study of multiple sclerosis in twins : update. *Ann. Neurol.*, 1993, **33** (3) : 281-285.
17. EBERS G. C., SADOVNICK A. D., RISCH N. J. A genetic basis for familial aggregation in multiple sclerosis. Canadian Collaborative Study Group. *Nature*, 1995, **377** (6545) : 150-151.
18. SADOVNICK A. D., EBERS G. C., DYMENT D. A., RISCH N. J. Evidence for genetic basis of multiple sclerosis. The Canadian Collaborative Study Group. *Lancet*, 1996, **347** (9017) : 1728-1730.
19. BECK S., CANN H. M., CAMPBELL R. D., DUNHAM I., INOKO H., JAZWINSKA E. C., RAGOISSIS J., TROWSDALE J., ZIEGLER A. Third single chromosome 6 workshop : meeting report. *DNA Seq.*, 1997, **8** (3) : 113-129.
20. FOGDELL A., HILLERT J., SACHS C., OLERUP O. The multiple sclerosis- and narcolepsy-associated HLA class II haplotype includes the DRB5*0101 allele. *Tissue Antigens*, 1995, **46** (4) : 333-336.
21. FOGDELL-HAHN A., LIGERS A., GRONNING M., HILLERT J., OLERUP O. Multiple sclerosis : a modifying influence of HLA class I genes in an HLA class II associated autoimmune disease. *Tissue Antigens*, 2000, **55** (2) : 140-148.
22. DYMENT D. A., EBERS G. C., SADOVNICK A. D. Genetics of multiple sclerosis. *Lancet Neurol.*, 2004, **3** (2) : 104-110.
23. MCFARLAND H. F. Twin studies and multiple sclerosis. *Ann. Neurol.*, 1992, **32** (6) : 722-723.

24. KURTZKE J. F. Epidemiologic evidence for multiple sclerosis as an infection. *Clin. Microbiol. Rev.*, 1993, **6** (4) : 382-427.
25. DEAN G., KURTZKE J. F. On the risk of multiple sclerosis according to age at immigration to South Africa. *Br. Med. J.*, 1971, **3** (777) : 725-729.
26. COO H., ARONSON K. J. A systematic review of several potential non-genetic risk factors for multiple sclerosis. *Neuroepidemiology*, 2004, **23** (1-2) : 1-12.
27. JOHNSON R. T. The virology of demyelinating diseases. *Ann. Neurol.*, 1994, **36** Suppl : S54-S60.
28. SOLDAN S. S., JACOBSON S. Role of viruses in etiology and pathogenesis of multiple sclerosis. *Adv. Virus. Res.*, 2001, **56** : 517-555.
29. ALLEN I., BRANKIN B. Pathogenesis of multiple sclerosis – the immune diathesis and the role of viruses. *J. Neuropathol. Exp. Neurol.*, 1993, **52** (2) : 95-105.
30. MILLER S. D., VANDERLUGT C. L., BEGOLKA W. S., PAO W., YAUCH R. L., NEVILLE K. L., KATZ-LEVY Y., CARRIZOSA A., KIM B. S. Persistent infection with Theiler's virus leads to CNS autoimmunity via epitope spreading. *Nat. Med.*, 1997, **3** (10) : 1133-1136.
31. STOHLMAN S. A., HINTON D. R. Viral induced demyelination. *Brain Pathol.*, 2001, **11** (1) : 92-106.
32. WUCHERPFENNIG K. W., STROMINGER J. L. Molecular mimicry in T cell-mediated autoimmunity : viral peptides activate human T cell clones specific for myelin basic protein. *Cell*, 1995, **80** (5) : 695-705.
33. SOTGIU S., PUGLIATTI M., FOIS M. L., ARRU G., SANNA A., SOTGIU M. A., ROSATI G. Genes, environment, and susceptibility to multiple sclerosis. *Neurobiol. Dis.*, 2004, **17** (2) : 131-143.
34. KELLY M. A., CAVAN D. A., PENNY M. A., MIJOVIC C. H., JENKINS D., MORRISSEY S., MILLER D. H., BARNETT A. H., FRANCIS D. A. The influence of HLA-DR and -DQ alleles on progression to multiple sclerosis following a clinically isolated syndrome. *Hum. Immunol.*, 1993, **37** (3) : 185-191.
35. OTA K., MATSUI M., MILFORD E. L., MACKIN G. A., WEINER H. L., HAFLER D. A. T-cell recognition of an immunodominant myelin basic protein epitope in multiple sclerosis. *Nature*, 1990, **346** (6280) : 183-187.
36. ZAMVIL S. S., STEINMAN L. The T lymphocyte in experimental allergic encephalomyelitis. *Annu. Rev. Immunol.*, 1990, **8** : 579-621.
37. MARTIN R., MCFARLAND H. F., MCFARLIN D. E. Immunological aspects of demyelinating diseases. *Annu. Rev. Immunol.*, 1992, **10** : 153-187.
38. ZAMVIL S., NELSON P., TROTTER J., MITCHELL D., KNOBLER R., FRITZ R., STEINMAN L. T-cell clones specific for myelin basic protein induce chronic relapsing paralysis and demyelination. *Nature*, 1985, **317** (6035) : 355-358.
39. ZHANG J., MARKOVIC-PLESE S., LACET B., RAUS J., WEINER H. L., HAFLER D. A. Increased frequency of interleukin 2-responsive T cells specific for myelin basic protein and proteolipid protein in peripheral blood and cerebrospinal fluid of patients with multiple sclerosis. *J. Exp. Med.*, 1994, **179** (3) : 973-984.
40. OKSENBERG J. R., PANZARA M. A., BEGOVICH A. B., MITCHELL D., ERLICH H. A., MURRAY R. S., SHIMONKEVITZ R., SHERRITT M., ROTHBARD J., BERNARD C. C. Selection for T-cell receptor V beta-D beta-J beta gene rearrangements with specificity for a myelin basic protein peptide in brain lesions of multiple sclerosis. *Nature*, 1993, **362** (6415) : 68-70.
41. WUCHERPFENNIG K. W., SETTE A., SOUTHWOOD S., OSEROFF C., MATSUI M., STROMINGER J. L., HAFLER D. A. Structural requirements for binding of an immunodominant myelin basic protein peptide to DR2 isotypes and for its recognition by human T cell clones. *J. Exp. Med.*, 1994, **179** (1) : 279-290.
42. CHOU Y. K., VAINIENE M., WHITHAM R., BOURDETTE D., CHOU C. H., HASHIM G., OFFNER H., VANDENBARK A. A. Response of human T lymphocyte lines to myelin basic protein : association of dominant epitopes with HLA class II restriction molecules. *J. Neurosci. Res.*, 1989, **23** (2) : 207-216.
43. MARTIN R., HOWELL M. D., JARAQUEMADA D., FLERLAGE M., RICHERT J., BROSTOFF S., LONG E. O., MCFARLIN D. E., MCFARLAND H. F. A myelin basic protein peptide is recognized by cytotoxic T cells in the context of four HLA-DR types associated with multiple sclerosis. *J. Exp. Med.*, 1991, **173** (1) : 19-24.
44. FULLER K. G., OLSON J. K., HOWARD L. M., CROXFORD J. L., MILLER S. D. Mouse models of multiple sclerosis : experimental autoimmune encephalomyelitis and Theiler's virus-induced demyelinating disease. *Methods Mol. Med.*, 2004, **102** : 339-361.
45. FUJINAMI R. S., OLDSTONE M. B. Amino acid homology between the encephalitogenic site of myelin basic protein and virus : mechanism for autoimmunity. *Science*, 1985, **230** (4729) : 1043-1045.
46. CROXFORD J. L., OLSON J. K., MILLER S. D. Epitope spreading and molecular mimicry as triggers of autoimmunity in the Theiler's virus-induced demyelinating disease model of multiple sclerosis. *Autoimmun. Rev.*, 2002, **1** (5) : 251-260.
47. HEMMER B., JACOBSEN M., SOMMER N. Degeneracy in T-cell antigen recognition - implications for the pathogenesis of autoimmune diseases. *J. Neuroimmunol.*, 2000, **107** (2) : 148-153.
48. BROCKE S., GAUR A., PIERCY C., GAUTAM A., GIJBELS K., FATHMAN C. G., STEINMAN L. Induction of relapsing paralysis in experimental autoimmune encephalomyelitis by bacterial superantigen. *Nature*, 1993, **365** (6447) : 642-644.
49. WALDNER H., COLLINS M., KUCHROO V. K. Activation of antigen-presenting cells by microbial products breaks self tolerance and induces autoimmune disease. *J. Clin. Invest.*, 2004, **113** (7) : 990-997.
50. SEGAL B. M., DWYER B. K., SHEVACH E. M. An interleukin (IL)-10/IL-12 immunoregulatory circuit controls susceptibility to autoimmune disease. *J. Exp. Med.*, 1998, **187** (4) : 537-546.
51. OLSON J. K., CROXFORD J. L., MILLER S. D. Virus-induced autoimmunity : potential role of viruses in

- initiation, perpetuation, and progression of T-cell-mediated autoimmune disease. *Viral Immunol.*, 2001, **14** (3) : 227-250.
52. McMAHON E. J., BAILEY S. L., CASTENADA C. V., WALDNER H., MILLER S. D. Epitope spreading initiates in the CNS in two mouse models of multiple sclerosis. *Nat. Med.*, 2005, **11** (3) : 335-339.
 53. GRETER M., HEPPNER F. L., LEMOS M. P., ODERMATT B. M., GOEBELS N., LAUFER T., NOELLE R. J., BECHER B. Dendritic cells permit immune invasion of the CNS in an animal model of multiple sclerosis. *Nat. Med.*, 2005, **11** (3) : 328-334.
 54. KRAKOWSKI M. L., OWENS T. Naive T lymphocytes traffic to inflamed central nervous system, but require antigen recognition for activation. *Eur. J. Immunol.*, 2000, **30** (4) : 1002-1009.
 55. CANNELLA B., CROSS A. H., RAINE C. S. Adhesion-related molecules in the central nervous system. Upregulation correlates with inflammatory cell influx during relapsing experimental autoimmune encephalomyelitis. *Lab. Invest.*, 1991, **65** (1) : 23-31.
 56. HAFLER D. A., WEINER H. L. In vivo labeling of blood T cells : rapid traffic into cerebrospinal fluid in multiple sclerosis. *Ann. Neurol.*, 1987, **22** (1) : 89-93.
 57. STEINMAN L., MARTIN R., BERNARD C., CONLON P., OKSENBERG J. R. Multiple sclerosis : deeper understanding of its pathogenesis reveals new targets for therapy. *Annu. Rev. Neurosci.*, 2002, **25** : 491-505.
 58. PERSIDSKY Y. Model systems for studies of leukocyte migration across the blood - brain barrier. *J. Neurovirol.*, 1999, **5** (6) : 579-590.
 59. BUTCHER E. C., PICKER L. J. Lymphocyte homing and homeostasis. *Science*, 1996, **272** (5258) : 60-66.
 60. HARTUNG H. P., ARCHELOS J. J., ZIELASEK J., GOLD R., KOLTZENBURG M., REINERS K. H., TOYKA K. V. Circulating adhesion molecules and inflammatory mediators in demyelination : a review. *Neurology*, 1995, **45** (6 Suppl. 6) : S22-S32.
 61. CUZNER M. L., OPDENAKKER G. Plasminogen activators and matrix metalloproteases, mediators of extracellular proteolysis in inflammatory demyelination of the central nervous system. *J. Neuroimmunol.*, 1999, **94** (1-2) : 1-14.
 62. YONG V. W., KREKOSKI C. A., FORSYTH P. A., BELL R., EDWARDS D. R. Matrix metalloproteinases and diseases of the CNS. *Trends Neurosci.*, 1998, **21** (2) : 75-80.
 63. LEPPERT D., WAUBANT E., GALARDY R., BUNNETT N. W., HAUSER S. L. T cell gelatinases mediate basement membrane transmigration in vitro. *J. Immunol.*, 1995, **154** (9) : 4379-4389.
 64. FIERZ W., ENDLER B., RESKE K., WEKERLE H., FONTANA A. Astrocytes as antigen-presenting cells. I. Induction of Ia antigen expression on astrocytes by T cells via immune interferon and its effect on antigen presentation. *J. Immunol.*, 1985, **134** (6) : 3785-3793.
 65. CARSON M. J., REILLY C. R., SUTCLIFFE J. G., LO D. Mature microglia resemble immature antigen-presenting cells. *Glia*, 1998, **22** (1) : 72-85.
 66. HICKEY W. F., KIMURA H. Perivascular microglial cells of the CNS are bone marrow-derived and present antigen in vivo. *Science*, 1988, **239** (4837) : 290-292.
 67. BECHER B., ANTEL J. P. Comparison of phenotypic and functional properties of immediately ex vivo and cultured human adult microglia. *Glia*, 1996, **18** (1) : 1-10.
 68. BROSNAN C. F., RAINE C. S. Mechanisms of immune injury in multiple sclerosis. *Brain Pathol.*, 1996, **6** (3) : 243-257.
 69. HELLINGS N., RAUS J., STINISSEN P. Insights into the immunopathogenesis of multiple sclerosis. *Immunol. Res.*, 2002, **25** (1) : 27-51.
 70. COMPSTON A. Remyelination of the central nervous system. *Mult. Scler.*, 1996, **1** (6) : 388-392.
 71. PRINEAS J. W., BARNARD R. O., KWON E. E., SHARER L. R., CHO E. S. Multiple sclerosis : remyelination of nascent lesions. *Ann. Neurol.*, 1993, **33** (2) : 137-151.
 72. RAINE C. S., WU E. Multiple sclerosis : remyelination in acute lesions. *J. Neuropathol. Exp. Neurol.*, 1993, **52** (3) : 199-204.
 73. SCOLDING N., FRANKLIN R., STEVENS S., HELDIN C. H., COMPSTON A., NEWCOMBE J. Oligodendrocyte progenitors are present in the normal adult human CNS and in the lesions of multiple sclerosis. *Brain*, 1998, **121** (Pt 12) : 2221-2228.
 74. PERIER O., GREGOIRE A. Electron microscopic features of multiple sclerosis lesions. *Brain*, 1965, **88** (5) : 937-952.
 75. SUZUKI K., ANDREWS J. M., WALTZ J. M., TERRY R. D. Ultrastructural studies of multiple sclerosis. *Lab. Invest.*, 1969, **20** (5) : 444-454.
 76. ZHAO C., FANCY S. P., KOTTER M. R., LI W. W., FRANKLIN R. J. Mechanisms of CNS remyelination - the key to therapeutic advances. *J. Neurol. Sci.*, 2005, **233** (1-2) : 87-91.
 77. LUCCHINETTI C. F., BRUECK W., RODRIGUEZ M., LASSMANN H. Multiple sclerosis : lessons from neuropathology. *Semin. Neurol.*, 1998, **18** (3) : 337-349.
 78. KORNEK B., LASSMANN H. Neuropathology of multiple sclerosis-new concepts. *Brain Res. Bull.*, 2003, **61** (3) : 321-326.
 79. BRUCK W., BITSCH A., KOLENDA H., BRUCK Y., STIEFEL M., LASSMANN H. Inflammatory central nervous system demyelination : correlation of magnetic resonance imaging findings with lesion pathology. *Ann. Neurol.*, 1997, **42** (5) : 783-793.
 80. BITSCH A., SCHUCHARDT J., BUNKOWSKI S., KUHLMANN T., BRUCK W. Acute axonal injury in multiple sclerosis. Correlation with demyelination and inflammation. *Brain*, 2000, **123** (Pt 6) : 1174-1183.
 81. REINDL M., LININGTON C., BREHM U., EGG R., DILITZ E., DEISENHAMMER F., POEWE W., BERGER T. Antibodies against the myelin oligodendrocyte glycoprotein and the myelin basic protein in multiple sclerosis and other neurological diseases : a comparative study. *Brain*, 1999, **122** (Pt 11) : 2047-2056.
 82. KORNEK B., LASSMANN H. Axonal pathology in multiple sclerosis. A historical note. *Brain Pathol.*, 1999, **9** (4) : 651-656.

83. FERGUSON B., MATYSZAK M. K., ESIRI M. M., PERRY V. H. Axonal damage in acute multiple sclerosis lesions. *Brain*, 1997, **120** (Pt 3) : 393-399.
84. TRAPP B. D., PETERSON J., RANSOHOFF R. M., RUDICK R., MORK S., BO L. Axonal transection in the lesions of multiple sclerosis. *N. Engl. J. Med.*, 1998, **338** (5) : 278-285.
85. KORNEK B., STORCH M. K., WEISSERT R., WALLSTROEM E., STEFFERL A., OLSSON T., LININGTON C., SCHMIDBAUER M., LASSMANN H. Multiple sclerosis and chronic autoimmune encephalomyelitis : a comparative quantitative study of axonal injury in active, inactive, and remyelinated lesions. *Am. J. Pathol.*, 2000, **157** (1) : 267-276.
86. KUHLMANN T., LINGFELD G., BITSCH A., SCHUCHARDT J., BRUCK W. Acute axonal damage in multiple sclerosis is most extensive in early disease stages and decreases over time. *Brain*, 2002, **125** (Pt 10) : 2202-2212.
87. BJARTMAR C., WUJEK J. R., TRAPP B. D. Axonal loss in the pathology of MS : consequences for understanding the progressive phase of the disease. *J. Neurol. Sci.*, 2003, **206** (2) : 165-171.
88. TRAPP B. D., RANSOHOFF R., RUDICK R. Axonal pathology in multiple sclerosis : relationship to neurologic disability. *Curr. Opin. Neurol.*, 1999, **12** (3) : 295-302.
89. BJARTMAR C., TRAPP B. D. Axonal and neuronal degeneration in multiple sclerosis : mechanisms and functional consequences. *Current Opinion in Neurology*, 2001, **14** (3) : 271-278.
90. MATTHEWS P. M., DE STEFANO N., NARAYANAN S., FRANCIS G. S., WOLINSKY J. S., ANTEL J. P., ARNOLD D. L. Putting magnetic resonance spectroscopy studies in context : axonal damage and disability in multiple sclerosis. *Semin. Neurol.*, 1998, **18** (3) : 327-336.
91. HAFNER D. A. Multiple sclerosis. *J. Clin. Invest.*, 2004, **113** (6) : 788-794.
92. PETTINELLI C. B., MCFARLIN D. E. Adoptive transfer of experimental allergic encephalomyelitis in SJL/J mice after in vitro activation of lymph node cells by myelin basic protein : requirement for Lyt 1+ 2- T lymphocytes. *J. Immunol.*, 1981, **127** (4) : 1420-1423.
93. HUANG D., HAN Y., RANI M. R., GLABINSKI A., TREBST C., SORENSEN T., TANI M., WANG J., CHIEN P., O'BRYAN S., BIELECKI B., ZHOU Z. L., MAJUMDER S., RANSOHOFF R. M. Chemokines and chemokine receptors in inflammation of the nervous system : manifold roles and exquisite regulation. *Immunol. Rev.*, 2000, **177** : 52-67.
94. MISU T., ONODERA H., FUJIHARA K., MATSUSHIMA K., YOSHIE O., OKITA N., TAKASE S., ITOYAMA Y. Chemokine receptor expression on T cells in blood and cerebrospinal fluid at relapse and remission of multiple sclerosis : imbalance of Th1/Th2-associated chemokine signaling. *J. Neuroimmunol.*, 2001, **114** (1-2) : 207-212.
95. GIUNTI D., BORSSELLINO G., BENELLI R., MARCHESI M., CAPELLO E., VALLE M. T., PEDEMONTE E., NOONAN D., ALBINI A., BERNARDI G., MANCARDI G. L., BATTISTINI L., UCCELLI A. Phenotypic and functional analysis of T cells homing into the CSF of subjects with inflammatory diseases of the CNS. *J. Leukoc. Biol.*, 2003, **73** (5) : 584-590.
96. BABBE H., ROERS A., WAISMAN A., LASSMANN H., GOEBELS N., HOHLFELD R., FRIESE M., SCHRODER R., DECKERT M., SCHMIDT S., RAVID R., RAJEWSKY K. Clonal expansions of CD8 (+) T cells dominate the T cell infiltrate in active multiple sclerosis lesions as shown by micromanipulation and single cell polymerase chain reaction. *J. Exp. Med.*, 2000, **192** (3) : 393-404.
97. GAY F. W., DRYE T. J., DICK G. W., ESIRI M. M. The application of multifactorial cluster analysis in the staging of plaques in early multiple sclerosis. Identification and characterization of the primary demyelinating lesion. *Brain*, 1997, **120** (Pt 8) : 1461-1483.
98. JACOBSEN M., CEPOK S., QUAK E., HAPPEL M., GABER R., ZIEGLER A., SCHOCK S., OERTEL W. H., SOMMER N., HEMMER B. Oligoclonal expansion of memory CD8+ T cells in cerebrospinal fluid from multiple sclerosis patients. *Brain*, 2002, **125** (Pt 3) : 538-550.
99. SKULINA C., SCHMIDT S., DORNMAIR K., BABBE H., ROERS A., RAJEWSKY K., WEKERLE H., HOHLFELD R., GOEBELS N. Multiple sclerosis : brain-infiltrating CD8+ T cells persist as clonal expansions in the cerebrospinal fluid and blood. *Proc. Natl. Acad. Sci. USA*, 2004, **101** (8) : 2428-2433.
100. JUREWICZ A., BIDDISON W. E., ANTEL J. P. MHC class I-restricted lysis of human oligodendrocytes by myelin basic protein peptide-specific CD8 T lymphocytes. *Journal of Immunology*, 1998, **160** (6) : 3056-3059.
101. MEDANA I. M., GALLIMORE A., OXENIUS A., MARTINIC M. M., WEKERLE H., NEUMANN H. MHC class I-restricted killing of neurons by virus-specific CD8+ T lymphocytes is effected through the Fas/FasL, but not the perforin pathway. *Eur. J. Immunol.*, 2000, **30** (12) : 3623-3633.
102. CRAWFORD M. P., YAN S. X., ORTEGA S. B., MEHTA R. S., HEWITT R. E., PRICE D. A., STASTNY P., DOUEK D. C., KOUP R. A., RACKE M. K., KARANDIKAR N. J. High prevalence of autoreactive, neuroantigen-specific CD8+ T cells in multiple sclerosis revealed by novel flow cytometric assay. *Blood*, 2004, **103** (11) : 4222-4231.
103. TSUCHIDA T., PARKER K. C., TURNER R. V., MCFARLAND H. F., COLIGAN J. E., BIDDISON W. E. Autoreactive CD8+ T-cell responses to human myelin protein-derived peptides. *Proc. Natl. Acad. Sci. USA*, 1994, **91** (23) : 10859-10863.
104. HONMA K., PARKER K. C., BECKER K. G., MCFARLAND H. F., COLIGAN J. E., BIDDISON W. E. Identification of an epitope derived from human proteolipid protein that can induce autoreactive CD8+ cytotoxic T lymphocytes restricted by HLA-A3 : evidence for cross-reactivity with an environmental microorganism. *J. Neuroimmunol.*, 1997, **73** (1-2) : 7-14.
105. ZANG Y. C., LI S., RIVERA V. M., HONG J., ROBINSON R. R., BREITBACH W. T., KILLIAN J., ZHANG J. Z. Increased CD8+ cytotoxic T cell responses to myelin basic protein in multiple sclerosis. *J. Immunol.*, 2004, **172** (8) : 5120-5127.

106. HUSEBY E. S., LIGGITT D., BRABB T., SCHNABEL B., OHLEN C., GOVERMAN J. A pathogenic role for myelin-specific CD8 (+) T cells in a model for multiple sclerosis. *J. Exp. Med.*, 2001, **194** (5) : 669-676.
107. STEINMAN L. Myelin-specific CD8 T cells in the pathogenesis of experimental allergic encephalitis and multiple sclerosis. *J. Exp. Med.*, 2001, **194** (5) : F27-F30.
108. DRESSEL A., CHIN J. L., SETTE A., GAUSLING R., HOLLSBERG P., HAFLER D. A. Autoantigen recognition by human CD8 T cell clones : enhanced agonist response induced by altered peptide ligands. *J. Immunol.*, 1997, **159** (10) : 4943-4951.
109. NEUMANN H., MEDANA I. M., BAUER J., LASSMANN H. Cytotoxic T lymphocytes in autoimmune and degenerative CNS diseases. *Trends Neurosci.*, 2002, **25** (6) : 313-319.
110. GIULIANI F., GOODYER C. G., ANTEL J. P., YONG V. W. Vulnerability of human neurons to T cell-mediated cytotoxicity. *J. Immunol.*, 2003, **171** (1) : 368-379.
111. HELTINGS N., RAUS J., STINISSEN P. T-cell vaccination in multiple sclerosis : update on clinical application and mode of action. *Autoimmun. Rev.*, 2004, **3** (4) : 267-275.
112. FERNANDEZ O., ARBIZU T., IZQUIERDO G., MARTINEZ-YELAMOS A., GATA J. M., LUQUE G., DE RAMON E. Clinical benefits of interferon beta-1a in relapsing-remitting MS : a phase IV study. *Acta Neurol. Scand.*, 2003, **107** (1) : 7-11.
113. LEARY S. M., THOMPSON A. J. Interferon beta-1a in primary progressive multiple sclerosis. *J. Neurol. Sci.*, 2003, **206** (2) : 215-216.
114. PATY D. W., LI D. K. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. II. MRI analysis results of a multicenter, randomized, double-blind, placebo-controlled trial., 1993 [classical article]. *Neurology*, 2001, **57** (12 Suppl. 5) : S10-S15.
115. PATY D. W., LI D. K. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. II. MRI analysis results of a multicenter, randomized, double-blind, placebo-controlled trial. UBC MS/MRI Study Group and the IFNB Multiple Sclerosis Study Group. *Neurology*, 1993, **43** (4) : 662-667.
116. YONG V. W., CHABOT S., STUVE O., WILLIAMS G. Interferon beta in the treatment of multiple sclerosis : mechanisms of action. *Neurology*, 1998, **51** (3) : 682-689.
117. DHIB-JALBUT S. Mechanisms of action of interferons and glatiramer acetate in multiple sclerosis. *Neurology*, 2002, **58** (8 Suppl. 4) : S3-S9.
118. NEUHAUS O., FARINA C., WEKERLE H., HOHLFELD R. Mechanisms of action of glatiramer acetate in multiple sclerosis. *Neurology*, 2001, **56** (6) : 702-708.
119. ZIEMSEN T., KUMPFEL T., KLINKERT W. E., NEUHAUS O., HOHLFELD R. Glatiramer acetate-specific T-helper 1- and 2-type cell lines produce BDNF : implications for multiple sclerosis therapy. Brain-derived neurotrophic factor. *Brain*, 2002, **125** (Pt 11) : 2381-2391.
120. CHEN M., VALENZUELA R. M., DHIB-JALBUT S. Glatiramer acetate-reactive T cells produce brain-derived neurotrophic factor. *J. Neurol. Sci.*, 2003, **215** (1-2) : 37-44.
121. THE IFNB MULTIPLE SCLEROSIS STUDY GROUP. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. I. Clinical results of a multicenter, randomized, double-blind, placebo-controlled trial. The IFNB Multiple Sclerosis Study Group. *Neurology*, 1993, **43** (4) : 655-661.
122. THE IFNB MULTIPLE SCLEROSIS STUDY GROUP AND THE UNIVERSITY OF BRITISH COLUMBIA MS/MRI ANALYSIS GROUP. Interferon beta-1b in the treatment of multiple sclerosis : final outcome of the randomized controlled trial. The IFNB Multiple Sclerosis Study Group and The University of British Columbia MS/MRI Analysis Group. *Neurology*, 1995, **45** (7) : 1277-1285.
123. JOHNSON K. P., BROOKS B. R., COHEN J. A., FORD C. C., GOLDSTEIN J., LISAK R. P., MYERS L. W., PANITCH H. S., ROSE J. W., SCHIFFER R. B. Copolymer 1 reduces relapse rate and improves disability in relapsing-remitting multiple sclerosis : results of a phase III multicenter, double-blind placebo-controlled trial. The Copolymer 1 Multiple Sclerosis Study Group. *Neurology*, 1995, **45** (7) : 1268-1276.
124. FILIPPINI G., MUNARI L., INCORVAIA B., EBERS G. C., POLMAN C., D'AMICO R., RICE G. P. Interferons in relapsing remitting multiple sclerosis : a systematic review. *Lancet*, 2003, **361** (9357) : 545-552.
125. COHEN J. A., CUTTER G. R., FISCHER J. S., GOODMAN A. D., HEIDENREICH F. R., KOOLJMAN M. F., SANDROCK A. W., RUDICK R. A., SIMON J. H., SIMONIAN N. A., TSAO E. C., WHITAKER J. N. Benefit of interferon beta-1a on MSFC progression in secondary progressive MS. *Neurology*, 2002, **59** (5) : 679-687.
126. POLMAN C. H., UITDEHAAG B. M. New and emerging treatment options for multiple sclerosis. *Lancet Neurol.*, 2003, **2** (9) : 563-566.
127. JACOBS L. D., BECK R. W., SIMON J. H., KINKEL R. P., BROWNSCHIEDLE C. M., MURRAY T. J., SIMONIAN N. A., SLASOR P. J., SANDROCK A. W. Intramuscular interferon beta-1a therapy initiated during a first demyelinating event in multiple sclerosis. CHAMPS Study Group. *N. Engl. J. Med.*, 2000, **343** (13) : 898-904.
128. COMI G., FILIPPI M., BARKHOF F., DURELLI L., EDAN G., FERNANDEZ O., HARTUNG H., SEELDRAYERS P., SORENSEN P. S., ROVARIS M., MARTINELLI V., HOMMES O. R. Effect of early interferon treatment on conversion to definite multiple sclerosis : a randomised study. *Lancet*, 2001, **357** (9268) : 1576-1582.
129. HARTUNG H. P., GONSETTE R., KONIG N., KWIECINSKI H., GUSEO A., MORRISSEY S. P., KRAPP H., ZWINGERS T. Mitoxantrone in progressive multiple sclerosis : a placebo-controlled, double-blind, randomised, multicentre trial. *Lancet*, 2002, **360** (9350) : 2018-2025.
130. EDAN G., MILLER D., CLANET M., CONFAVREUX C., LYON-CAEN O., LUBETZKI C., BROCHET B., BERRY I., ROLLAND Y., FROMENT J. C., CABANIS E., IBAZIZEN M. T., GANDON J. M., LAI H. M., MOSELEY I., SABOURAUD O. Therapeutic effect of mitoxantrone combined with methylprednisolone in multiple

- sclerosis : a randomised multicentre study of active disease using MRI and clinical criteria. *J. Neurol. Neurosurg. Psychiatry*, 1997, **62** (2) : 112-118.
131. HOHLFELD R., WIENDL H. The ups and downs of multiple sclerosis therapeutics. *Ann. Neurol.*, 2001, **49** (3) : 281-284.
 132. HUGHES R. A. Immunotherapy for multiple sclerosis. *J. Neurol. Neurosurg. Psychiatry*, 1994, **57** (1) : 3-6.
 133. BARNES M. P., BATEMAN D. E., CLELAND P. G., DICK D. J., WALLS T. J., NEWMAN P. K., SAUNDERS M., TILLEY P. J. Intravenous methylprednisolone for multiple sclerosis in relapse. *J. Neurol. Neurosurg. Psychiatry*, 1985, **48** (2) : 157-159.
 134. MILLIGAN N. M., NEWCOMBE R., COMPSTON D. A. A double-blind controlled trial of high dose methylprednisolone in patients with multiple sclerosis : 1. Clinical effects. *J. Neurol. Neurosurg. Psychiatry*, 1987, **50** (5) : 511-516.
 135. BLEVINS G., MARTIN R. Future immunotherapies in multiple sclerosis. *Semin. Neurol.*, 2003, **23** (2) : 147-158.
 136. ZHANG J. T-cell vaccination in multiple sclerosis : immunoregulatory mechanism and prospects for therapy. *Crit. Rev. Immunol.*, 2001, **21** (1-3) : 41-55.
 137. VANDENBARK A. A., CHOU Y. K., WHITHAM R., MASS M., BUENAFE A., LIEFELD D., KAVANAGH D., COOPER S., HASHIM G. A., OFFNER H. Treatment of multiple sclerosis with T-cell receptor peptides : results of a double-blind pilot trial. *Nat. Med.*, 1996, **2** (10) : 1109-1115.
 138. BIELEKOVA B., RICHERT N., HOWARD T., BLEVINS G., MARKOVIC-PLESE S., MCCARTIN J., FRANK J. A., WURFEL J., OHAYON J., WALDMANN T. A., MCFARLAND H. F., MARTIN R. Humanized anti-CD25 (daclizumab) inhibits disease activity in multiple sclerosis patients failing to respond to interferon beta. *Proc. Natl. Acad. Sci. USA*, 2004, **101** (23) : 8705-8708.
 139. MILLER D. H., KHAN O. A., SHEREMATA W. A., BLUMHARDT L. D., RICE G. P., LIBONATI M. A., WILLMER-HULME A. J., DALTON C. M., MISZKIEL K. A., O'CONNOR P. W. A controlled trial of natalizumab for relapsing multiple sclerosis. *N. Engl. J. Med.*, 2003, **348** (1) : 15-23.
 140. WEBER F., POLAK T., GUNTHER A., KUBUSCHOK B., JANOVSKAJA J., BITSCH A., POSER S., RIECKMANN P. Synergistic immunomodulatory effects of interferon-beta1b and the phosphodiesterase inhibitor pentoxifylline in patients with relapsing-remitting multiple sclerosis. *Ann. Neurol.*, 1998, **44** (1) : 27-34.
 141. PAOLILLO A., COLES A. J., MOLYNEUX P. D., GAWNECAIN M., MACMANUS D., BARKER G. J., COMPSTON D. A., MILLER D. H. Quantitative MRI in patients with secondary progressive MS treated with monoclonal antibody Campath 1H. *Neurology*, 1999, **53** (4) : 751-757.
 142. YOUSSEF S., STUVE O., PATARROYO J. C., RUIZ P. J., RADOSEVICH J. L., HUR E. M., BRAVO M., MITCHELL D. J., SOBEL R. A., STEINMAN L., ZAMVIL S. S. The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature*, 2002, **420** (6911) : 78-84.
 143. PAHAN K., NAMBOODIRI A. M., SHEIKH F. G., SMITH B. T., SINGH I. Increasing cAMP attenuates induction of inducible nitric-oxide synthase in rat primary astrocytes. *J. Biol. Chem.*, 1997, **272** (12) : 7786-7791.
 144. WEITZ-SCHMIDT G., WELZENBACH K., BRINKMANN V., KAMATA T., KALLEN J., BRUNS C., COTTENS S., TAKADA Y., HOMMEL U. Statins selectively inhibit leukocyte function antigen-1 by binding to a novel regulatory integrin site. *Nat. Med.*, 2001, **7** (6) : 687-692.
 145. SICOTTE N. L., LIVA S. M., KLUTCH R., PFEIFFER P., BOUVIER S., ODESA S., WU T. C., VOSKUHL R. R. Treatment of multiple sclerosis with the pregnancy hormone estriol. *Ann. Neurol.*, 2002, **52** (4) : 421-428.
 146. DUNCAN I. D., GREVER W. E., ZHANG S. C. Repair of myelin disease : strategies and progress in animal models. *Mol. Med. Today*, 1997, **3** (12) : 554-561.
 147. FRANKLIN R. J. Remyelination of the demyelinated CNS : the case for and against transplantation of central, peripheral and olfactory glia. *Brain Res. Bull.*, 2002, **57** (6) : 827-832.
 148. BARON-VAN EVERCOOREN A., BLAKEMORE W. Remyelination through engraftment. In : *Myelin biology and disorders*. LAZZARINI R. (ed). San Diego : Elsevier, 2004 : 143-172.
 149. MOSCARELLO M. A., MAK B., NGUYEN T. A., WOOD D. D., MASTRONARDI F., LUDWIN S. K. Paclitaxel (Taxol) attenuates clinical disease in a spontaneously demyelinating transgenic mouse and induces remyelination. *Mult. Scler.*, 2002, **8** (2) : 130-138.
 150. CALABRESI P. A., WILTERDINK J. L., ROGG J. M., MILLS P., WEBB A., WHARTENBY K. A. An open-label trial of combination therapy with interferon beta-1a and oral methotrexate in MS. *Neurology*, 2002, **58** (2) : 314-317.
 151. SOOS J. M., STUVE O., YOUSSEF S., BRAVO M., JOHNSON H. M., WEINER H. L., ZAMVIL S. S. Cutting edge : oral type I IFN-tau promotes a Th2 bias and enhances suppression of autoimmune encephalomyelitis by oral glatiramer acetate. *J. Immunol.*, 2002, **169** (5) : 2231-2235.

Dr. N. HELLINGS,
 Biomedisch Onderzoeksinstituut,
 Hasselt University,
 Agoralaan building A,
 B-3590 Diepenbeek (Belgium).
 E-mail : niels.hellings@uhasselt.be.